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Ca²⁺-dependent nitric oxide release in the injured endothelium of excised rat aorta: a promising mechanism applying in vascular prosthetic devices in aging patients

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Abstract

Background: Nitric oxide is key to endothelial regeneration, but it is still unknown whether endothelial cell (EC) loss results in an increase in NO levels at the wound edge. We have already shown that endothelial damage induces a long-lasting Ca²⁺ entry into surviving cells through connexin hemichannels (CxHcs) uncoupled from their counterparts on ruptured cells. The physiological outcome of injury-induced Ca²⁺ inflow is, however, unknown.

Methods: In this study, we sought to determine whether and how endothelial scraping induces NO production (NOP) in the endothelium of excised rat aorta by exploiting the NO-sensitive fluorochrome, DAF-FM diacetate and the Ca²⁺-sensitive fluorescent dye, Fura-2/AM.

Results: We demonstrated that injury-induced NOP at the lesion site is prevented in presence of the endothelial NO synthase inhibitor, L-NAME, and in absence of extracellular Ca²⁺. Unlike ATP-dependent NO liberation, the NO response to injury is insensitive to BTP-2, which selectively blocks store-operated Ca²⁺ inflow. However, injury-induced NOP is significantly reduced by classic gap junction blockers, and by connexin mimetic peptides specifically targeting Cx37Hcs, Cx40Hcs, and Cx43Hcs. Moreover, disruption of caveolar integrity prevents injury-elicited NO signaling, but not the accompanying Ca²⁺ response.

Conclusions: The data presented provide the first evidence that endothelial scraping stimulates NO synthesis at the wound edge, which might both exert an immediate anti-thrombotic and anti-inflammatory action and promote the subsequent re-endothelialization.

Background

Endothelial injury is regarded as the early event that leads to the onset and progression of severe vascular disorders, such as thrombosis, hypertension, and atherosclerosis [1]. Endothelial cell (EC) loss physiologically occurs due to physiological turnover in focal areas of the inner surface of blood vessels which are adjacent to

regions with low or absent replication [2,3]. When the extent of EC loss is such small, i.e. limited to a belt of cells around the whole circumference of the arterial vessel, the re-endothelialization process is driven by proliferation, spreading, and migration of surviving ECs into the damaged site [4]. On the other hand, larger areas of injury may release chemical messages, such as concentration gradients of vascular endothelial growth factor (VEGF) and stromal derived factor-1 α (SDF-1 α), that recruit circulating endothelial progenitor cells to the wound edge in order to replace damaged endothelium [5-10]. The break

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in the anatomic integrity of vascular endothelium is significantly larger in subjects undergoing medical interventions, such as deployment of endovascular devices and percutaneous transluminal coronary angioplasty [2,11,12]. Such a heavy loss of ECs from the vascular wall dampens the beneficial effects of reconstructive surgery and prompts the quest for pharmacological treatments aiming at restoring the continuity of endothelial monolayer [2,11-13]. Accordingly, the de-endothelialisation of arterial vessels may cause thrombi formation and neointimal hyperplasia, giving raise to a process termed as “in-stent restenosis” (ISR) which renarrows the arterial lumen [2,11,12]. Drug-eluting stents (DES) may be implanted during or after angioplasty to inhibit neointimal hyperplasia and prevent ISR [2,12,14]. Unfortunately, the most commonly employed stents deliver drugs, such as sirolimus and paclitaxel, that cause a long-term inhibition of endothelial proliferation and migration. These untoward off-target effects significantly delay endothelial regrowth, thereby leaving uncovered the surface of the stent and increasing the risk of late in-stent thrombosis [14,15].

Aging is accompanied by a decline in the healthy function of multiple organ systems, leading to increased incidence of mortality from many diseases. Oxidative stress and elevated ROS (Reactive oxygen species) has been implicated in the mechanism of senescence and aging; they are also involved in cancer, diabetes, neurodegenerative, cardiovascular and other diseases [16,17]. Overproduction of oxidant molecules is due to several stress agents such chemicals, drugs, pollutants, high-caloric diets and exercise [18].

Nitric oxide (NO), a gasotransmitter that may be synthesized and released by ECs [1,19], might play a key role in healing injured endothelium. Accordingly, NO inhibits apoptosis and enhances EC proliferation, migration, and tubulogenesis [19]. Moreover, NO might serve as anti-inflammatory signal at the injured site by preventing local platelet activation and thrombus formation, by causing vasorelaxation, and by inhibiting the phenotypic switch of vascular smooth muscle cell (VSMC) [1,19]. Restoration of NO production at the site of vascular injury may attenuate neointimal hyperplasia and adverse the onset of the atherosclerotic process [20]. In vascular endothelium, NO may be synthesized by two different isoforms of NO synthase (NOS), namely inducible NOS (iNOS/NOS2), which mediates NO liberation during inflammatory reactions, and endothelial NOS (eNOS/NOS3), which is preferentially recruited by stimuli [19,21]. The catalytic reaction consists in the conversion of L-arginine to L-citrulline and requires several cofactors, such as NADPH, tetrahydrobiopterin and O_2 [19]. Unlike iNOS, which is inducible and Ca^{2+} -independent, both iNOS and eNOS are constitutive and bear a calmodulin-binding site

whose binding to cytosolic Ca^{2+} is essential to stimulate NO synthesis [19]. In several types of ECs, eNOS is preferentially recruited by Ca^{2+} store-operated Ca^{2+} entry (SOCE), which is gated by depletion of the inositol-1,4,5-trisphosphate ($InsP_3$)-sensitive stores within the endoplasmic reticulum (ER) [8,22]. This feature is due to the close proximity between eNOS and SOC channels at the caveolae, cholesterol-enriched surface microdomains that compartmentalize signal transduction molecules [23]. Cholesterol-binding drugs, such as methyl- β -cyclodextrin ($M\beta CD$), have been used to disrupt caveolae and impair NO signalling in vascular endothelium [24]. That NO is involved in EC response to injury has been suggested by the elevation in both eNOS protein and enzyme activity in the regenerating endothelium of rat aorta [25]. Moreover, a recent study carried out on this preparation reported an increase in $[Ca^{2+}]_i$ in ECs nearby the lesion site [2,9,26,27]. The Ca^{2+} response to injury comprises an initial peak, mainly due to Ca^{2+} release from $InsP_3$ -sensitive receptors ($InsP_3Rs$), which is followed by a long-lasting decay phase mediated by Ca^{2+} entry across the plasma membrane [27]. $InsP_3$ synthesis requires phospholipase C (PLC) activation by ATP (or ATP-derived nucleotides) released from ruptured cells [27]. Ca^{2+} inflow is supported by connexin (Cx) hemichannels (CxHcs), which have recently been described as alternative Ca^{2+} entry routes in vascular ECs as well as other non-excitable cell types [26-31]. Cxs may be classified on the basis of the molecular weight with three main isoforms (Cx37, Cx40, and Cx43) being found in ECs from several vascular beds, including rat aorta [32,33]. Despite the available evidences show an increase both in eNOS activity and in $[Ca^{2+}]_i$ within lesioned endothelium, there is no report of injury-induced NO release in blood vessels. Understanding the signal transduction machinery leading to endothelial activation and NO synthesis might be clinically relevant to design alternative strategies to restore endothelial integrity and prevent ISR.

In the present investigation, we sought to determine whether NO synthesis occurs in ECs facing the injury site of excised rat aorta. We further aimed at elucidating the signal transduction pathway responsible for NO release by lesioned endothelium. These goals were accomplished by loading ECs with Fura-2/AM, a Ca^{2+} -sensitive fluorochrome, and DAF-FM diacetate, a NO-sensitive fluorescent dye. We provided, for the first time, the evidence that mechanical injury induces a long lasting NO production (NOP) in cells nearby the lesion site which requires Ca^{2+} entry through uncoupled Cx37Hcs, Cx40Hcs, and Cx43Hc. These results might aid in developing novel pharmacological treatments devoted to restore endothelial integrity upon invasive medical interventions.

Methods

Dissection of the aorta

Wistar rats aged 2-3 months were sacrificed with an overdose of diethyl ether. The thoracic and abdominal aorta were dissected out and perfused with physiological salt solution (PSS). The vessel was cleaned of the surrounding connective tissue, cut in ~5 mm long rings, stored in PSS at room temperature (22-24 °C) and used within 5 hours. All the animal protocols were approved by the East Tennessee State University's Animal Care and Use Committee. All experiments conform to the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH Publication No. 85-23, revised 1996). The animal handling was under the continuous control of the Veterinary Surgeon of the University of Pavia.

Solutions

PSS had the following composition (in mM): 150 NaCl, 6 KCl, 1.5 CaCl₂, 1 MgCl₂, 10 Glucose, 10 Hepes. In Ca²⁺-free solution (0 Ca²⁺), Ca²⁺ was substituted with 2 mM NaCl, and 0.5 mM EGTA was added. Solutions were titrated to pH 7.4 with NaOH. Aortic rings were bathed in 0 Ca²⁺ for no longer than 90 sec before inducing the injury. Control experiments have demonstrated that such a short pre-incubation period is not able to deplete intracellular Ca²⁺ stores [27].

[Ca²⁺]_i and NO measurements.

The technique used to evaluate changes in [Ca²⁺]_i in intact endothelium has been previously described [26,27,34-37]. The aortic ring was opened and loaded with 16 μmol Fura-2/AM for 60 min at room temperature, washed and fixed by small pins with the luminal face up. *In situ* ECs were visualized by an upright epifluorescence Axiolab microscope (Carl Zeiss, Oberkochen, Germany), equipped with a Zeiss ×63 Achromplan objective (water-immersion, 2.0 mm working distance, 0.9 numerical aperture). ECs were excited alternately at 340 and 380 nm, and the emitted light was detected at 510 nm. The exciting filters were mounted on a filter wheel (Lambda 10, Sutter Instrument, Novato, CA, USA). Custom software, working in the LINUX environment, was used to drive the camera (Extended-ISIS Camera, Photonic Science, Millham, UK) and the filter wheel, and to measure and plot on-line the fluorescence from 10-15 rectangular "regions of interest" (ROI) enclosing one single cell. [Ca²⁺]_i was monitored by measuring, for each ROI, the ratio of the mean fluorescence emitted at 510 nm when exciting alternatively at 340 and 380 nm (shortly termed "ratio"). An increase in [Ca²⁺]_i causes an increase in the ratio. Ratio measurements were performed and plotted on-line every 5 s. The experiments were performed at room temperature (21-23 °C).

NO production in ECs in intact rat aorta was monitored with the membrane permeant 4-Amino-5-methylamino-2',7'-difluorofluorescein (DAF-FM) diacetate. Once open, the aortic ring was fixed by small pins with the luminal side facing up, loaded with 10 μM DAF-FM for 60 min at room temperature and washed in PSS for one hour. DAF-FM fluorescence was measured by using the same equipment described for Ca²⁺ recordings but with a different filter set, i.e. excitation at 480 nm and emission at 535 nm wavelength (emission intensity was shortly termed "NO_i"). NO measurements were performed and plotted on-line every 5 s. Again, off-line analysis was performed by using custom-made macros developed by Microsoft Office Excel software. The experiments were performed at room temperature. DAF-FM is essentially non-fluorescent until it irreversibly reacts with the nitrosonium cation produced by spontaneous oxidation of newly synthesized NO. The resulting fluorescent compound is trapped in the cytoplasm, so that DAF-FM fluorescence summates with continual NO production. As also found in bovine aortic ECs [38], DAF-FM fluorescence in rat aortic ECs underwent a persistent and linear increase likely due to the basal NO synthesis [19]. Accordingly, the change in baseline fluorescence was dramatically reduced when aortic rings were pre-incubated with L-NAME (5 mM), which inhibits eNOS by competing with its natural substrate, L-arginine (data not shown), or in 0Ca (data not shown). This observation was somehow expected since aortic endothelium has long been known to regulate vascular tone by basal release of NO [19]. Therefore, for each experiment, the basal DAF-FM signal was recorded for at least 20 minutes and the resulting curve fitted off-line by a linear regression equation in order to calculate the slope (i.e. the rate) of basal fluorescence increase. The slope was subtracted from the recorded trace, thus enabling to measure S-Nitroso-N-acetylpenicillamine (SNAP)-, ATP- and injury-induced NOP independently on basal eNOS activity [39] (see below). NO synthesis was evaluated by subtracting the baseline level (1 min before the increase in DAF-FM fluorescence) to the average value of the plateau phase (1 min before the end of the recording) recorded upon either ATP administration or endothelial scraping.

In order to assess the ability of the fluorimetric set up to detect evoked changes in intracellular NO levels, aortic rings were exposed to the NO donor, SNAP, and ATP, which has been shown to elicit NO production in a variety of ECs from different vascular districts [38,40]. As shown in Figure 1A and Figure 1B, respectively, SNAP (500 μM) and ATP (300 μM) caused an increase in the intensity of DAF-FM fluorescence in 19 out of 19 cells and in 170 out of 170 cells, respectively.

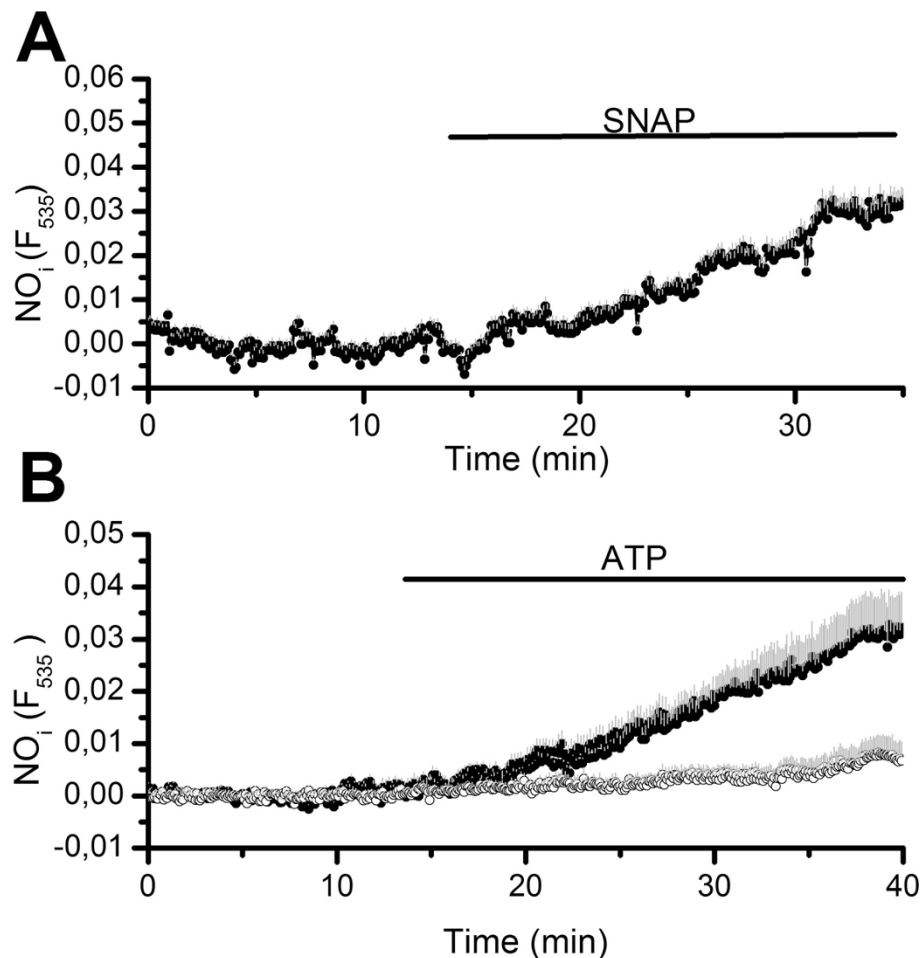


Figure 1 NO production in rat aortic endothelium. A, exposition to SNAP (500 μM) caused an increase in DAF-FM fluorescence in native rat aortic endothelium *in situ*. The trace is the mean \pm SE of 11 cells from the same visual field. B, ATP (300 μM) induced an elevation in intracellular NO (closed circles) that was dramatically hampered when the rings were pre-treated with L-NAME (5 mM) for 40 min (white circles). The traces are the mean \pm SE, respectively, of 10 (PSS) and 8 (L-NAME) cells from two aortic rings harvested from the same animal on the same day.

On average, L-NAME (5 mM) reduced by 86% ATP-induced NO production (0.025 ± 0.001 , $n = 170$, vs. 0.0041 ± 0.0009 , $n = 192$, $p < 0.001$) (Figure 1B).

Mechanical disruption of ECs

As shown in [26,27], aortic endothelium was injured under microscopic control by means of a glass microelectrode with a broken tip of about 30 μm diameter, driven by an XYZ hydraulic micromanipulator (Narishige, Japan). Images of either Fura-2 or DAF-FM/Fura-2 loaded ECs, together with numbered ROIs, were taken before the lesion, in order to identify the cells facing the injury site. As aforementioned, the dissection procedure could itself damage the intimal layer and cause SMCs to be loaded with Fura-2. These rings were, therefore, discarded. The microelectrode was first positioned almost parallel and very near to the endothelium surface. It was then moved downward, along the Z-axis, until the electrode tip gently touched the endothelium, and moved

horizontally across the visual field to scrape 1-3 consecutive rows of ECs along the whole diameter of the aortic ring. The following lesion was 170 μm long and 20 μm wide. This procedure, which allowed monitoring of NO signals in ECs adjacent to the lesion, mimics the ablation of the endothelial lining achieved in pre-clinical studies addressing the cellular mechanisms of intimal regrowth upon stent deployment [41-43]. The scraping of the endothelial monolayer imposed a physical stretching on the underlying SMCs which resembles the condition occurring during clinical interventions. Ethidium bromide (EB), a fluorescent molecule unable to cross an intact plasma membrane and therefore indicative of damaged/dead cells, was used to check the viability of both ECs nearby the injury and underlying SMCs [27]. When smooth muscle fibers were lesioned by mechanical scratching and stained by EB, the experiments were discarded.

Data analysis

For each protocol, data were collected from at least three rats. The amplitude of the peak Ca^{2+} response was measured as the difference between the ratio at the peak and the mean ratio of 1 min baseline before the peak. The details of NO measurements have been reported above. Only cells residing in the first row nearby the wound were used to elaborate the average value. For Ca^{2+} and NO measurements, statistical comparisons were made by Student's *t*-test for unpaired observations.

Chemicals

Fura-2/AM and DAF-FM was obtained from Molecular Probes (Molecular Probes Europe BV, Leiden, The Netherlands). N-(4-[3,5-bis(trifluoromethyl)-1H-pyrazol-1-yl]phenyl)-4-methyl-1,2,3-thiadiazole-5-carboxamide (BTP-2) was purchased from Calbiochem (La Jolla, CA, USA). All other chemicals were purchased from Sigma. Cx-mimetic peptides, including $^{37,43}\text{Gap27}$ and $^{40}\text{Gap27}$, were synthesized by Severn Biotech (Kidderminster, UK); purity was >95%. In more detail, $^{37,43}\text{Gap27}$ and $^{40}\text{Gap27}$ mimic a highly conserved SRPTEK sequence present in the second extracellular loop of Cxs 37 and 43 and of Cx 40, respectively [15,19]. Their scrambled versions were also synthesized by Severn Biotech. All other chemicals were of analytical grade and obtained from Sigma.

Results

Injury augments intracellular NO levels in the intact endothelium of rat aorta

Figure 2A depicts the injury-induced increase in $[\text{Ca}^{2+}]_i$ occurring within the ECs adjacent to the lesion site and which have lost their contact with the scraped cells. The Ca^{2+} signal consisted in an initial transient peak, mainly due to InsP_3 -dependent Ca^{2+} release, followed by a prolonged decay phase caused by Ca^{2+} entry from the extracellular milieu [27]. When the damage was performed on an aortic ring loaded with DAF-FM, it induced a gradual increase in fluorescence that reached a plateau after about 20 min (Figure 2B). This feature is consistent with the concomitant long-lasting Ca^{2+} inflow [27], which might sustain NO synthesis [19,38]. In the presence of the eNOS inhibitor, L-NAME (5 mM), mechanical damage failed to augment intracellular NO (Figure 2B). The statistical analysis is reported in Figure 3A. Similar to the intercellular Ca^{2+} wave, which did not spread farther than the 4th row of cells, the amplitude of the NO signal significantly ($p < 0.05$) decayed at this location (Figure 2C). This feature hints at a strong correlation between the increase in $[\text{Ca}^{2+}]_i$ triggered by the injury and the accompanying NO synthesis.

Injury-induced NOP depends on extracellular Ca^{2+} entry

It has long been known that, in vascular endothelium, eNOS is selectively activated by Ca^{2+} inflow rather than

intracellular Ca^{2+} release [8,22,40]. When mechanical damage was carried out in absence of extracellular Ca^{2+} , a maneuver which abolished the prolonged decay-phase of the Ca^{2+} signal, the injury did not result in a detectable NOP (Figure 3A). The involvement of intracellular Ca^{2+} mobilization was investigated with the aid of thapsigargin (2 μM), an inhibitor of the endoplasmic reticulum Ca^{2+} -ATPase, which prevents Ca^{2+} re-uptake into the stores and leads to their depletion. Preliminary experiments showed that pre-incubating the rings for 30 min with 2 μM thapsigargin abolished the intracellular Ca^{2+} mobilization triggered by supramaximal concentrations of ATP (300 μM) ($n = 79$; data not shown). Therefore, such a treatment causes the depletion of the Ca^{2+} reservoir underlying the initial Ca^{2+} response to injury [27]. The NO signal induced by endothelial scraping, however, was not significantly affected by thapsigargin (Figure 3A). Overall, these results strongly suggest that injury-promoted NOP in surviving ECs is sustained by Ca^{2+} entry, while is insensitive to Ca^{2+} release from ER. SOCE represents the preferential route for extracellular Ca^{2+} to engage eNOS in ECs [8,22,40]. However, BTP-2 (20 μM), a widely employed blocker of store-dependent Ca^{2+} inflow [6,8], does not significantly affect injury-induced NO synthesis (Figure 3A). We subsequently focused on the response to ATP, which elicits a BTP-2-sensitive SOCE in rat aortic endothelium [27]. ATP-induced NO synthesis was strongly inhibited in 0 Ca^{2+} (Figure 3B) and in the presence BTP-2 (Figure 3B). Collectively, these data concur with our previous findings on injury-induced Ca^{2+} inflow and rule out a detectable role for SOCE in NO release by lesioned endothelium.

Gap junction inhibitors reduce NO synthesis induced by Ca^{2+} entry in injured endothelium

The long-lasting decay phase of injury-induced Ca^{2+} elevation in rat aortic rings is dramatically diminished by gap junction blockers, such as octanol, palmitoleic acid and oleamide [26,27,44,45]. In agreement with the Ca^{2+} measurements [27], octanol (4 mM), oleamide (200 μM), and palmitoleic acid (50 μM) dramatically affected NO synthesis elicited by endothelial scraping (Figure 3C). In the light of our previous observations [27], these findings hint at a role for Ca^{2+} inflow through CxHcs in mediating NOP in injured endothelium. We assessed the specificity of gap junction blockers by measuring the Ca^{2+} response to ATP (20 μM) in the presence of octanol. At this dose, ATP evokes an InsP_3 -dependent Ca^{2+} release, which is subsequently buffered by the combined action of SERCA and $\text{Na}^+/\text{Ca}^{2+}$ exchanger [27,34,46]. As shown in Figure 4, pre-incubating rat aortic rings with 4 mM octanol for 20 min did not significantly affect either the magnitude of the $[\text{Ca}^{2+}]_i$ elevation or the rate of its decay phase to the baseline. This result concurs with previous evidence

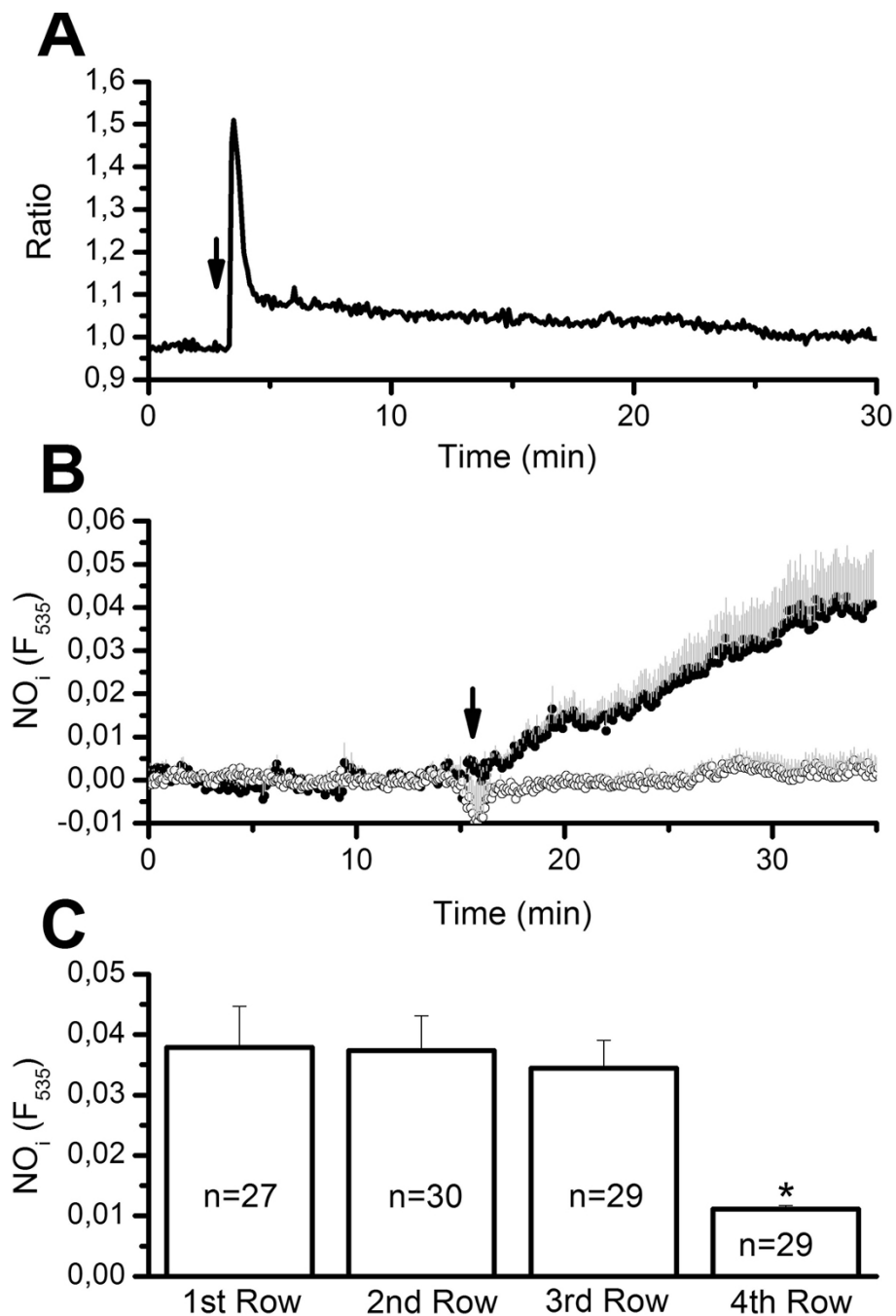


Figure 2 NO synthesis induced by mechanical injury rat aortic endothelium. A, Ca^{2+} signal elicited by lesioning rat aorta in PSS. The trace, recorded from a single cell, is representative of 10 cells from the same visual field. B, injury induced an elevation in intracellular NO levels that was abolished when the cells were pre-treated with the eNOS inhibitor, L-NAME (5 mM), for 40 min. The traces are the mean \pm SE, respectively, of 10 (PSS, closed circles) and 5 (L-NAME, open circles) cells from two aortic rings harvested from the same animal on the same day. C, mean \pm SE of the NO signal recorded at increasing distance from the lesion site. In this and the following figures, the arrow indicates when injury is performed. Asterisk indicates a level of significant difference <0.05.

about the efficacy of gap junction blockers as selective uncoupling drugs in rat aortic rings [47,48], and further support the role of CxHcs in mediating NO synthesis in injured endothelium.

Effect of Cx-mimetic peptides on injury-induced Ca^{2+} elevation and NO production

Rat aortic endothelium may express three Cx isoforms, namely Cx37, Cx40, and Cx43 [32,49], albeit a recent

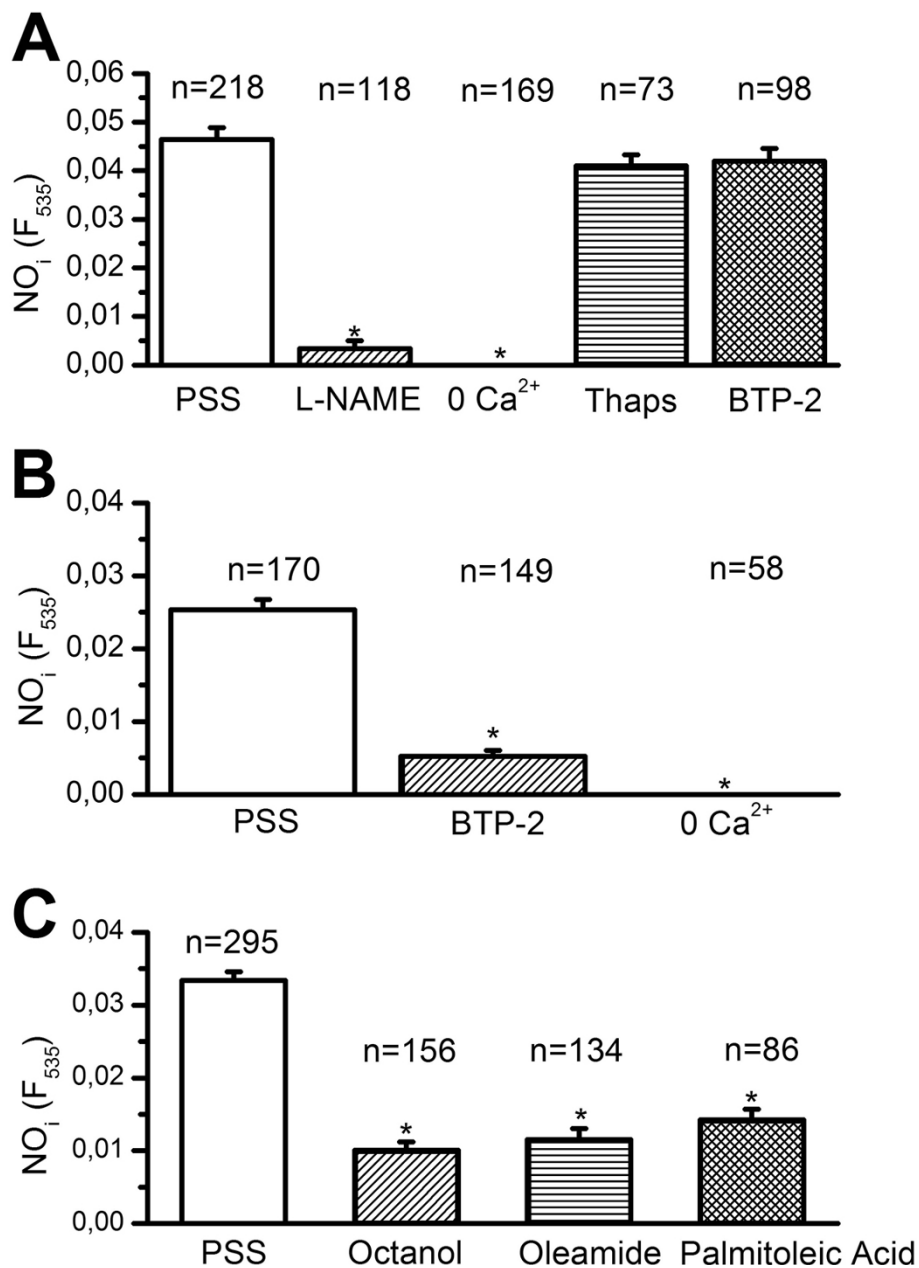


Figure 3 Amplitude of the NO response to injury (A and C) and ATP (B) following the designated treatments. The asterisk indicates a level of significant difference <0.05 . The number of ECs analyzed in each condition, harvested from 3-9 animals, is indicated above the bars. See the text for drug concentrations.

work reported the presence of Cx32 in various cultured human ECs [50]. The contribution of each Cx isotype to injury-evoked NO liberation was examined by exploiting Cx-mimetic peptides containing amino acid sequences corresponding to the Gap27 extracellular domain of Cx37 and Cx43 (^{37,43}Gap27) and to the Gap27 extracellular domain of Cx40 (⁴⁰Gap27). Although initially designed to impair gap junctional communication, the primary target of these compounds is provided by

CxHcs. The extracellular-loop sequence that interacts with the peptides is freely available in the CxHc form [51], so that full blockade of the unpaired hemichannel requires a much shorter pre-incubation (minutes vs. hours or longer) and much lower doses (micromolar vs. millimolar range) than inhibition of gap junctional coupling [51]. These compounds have been found to selectively inhibit Cx-dependent signalling in a variety of cell types, including rat aortic ECs [49,52], when the genetic

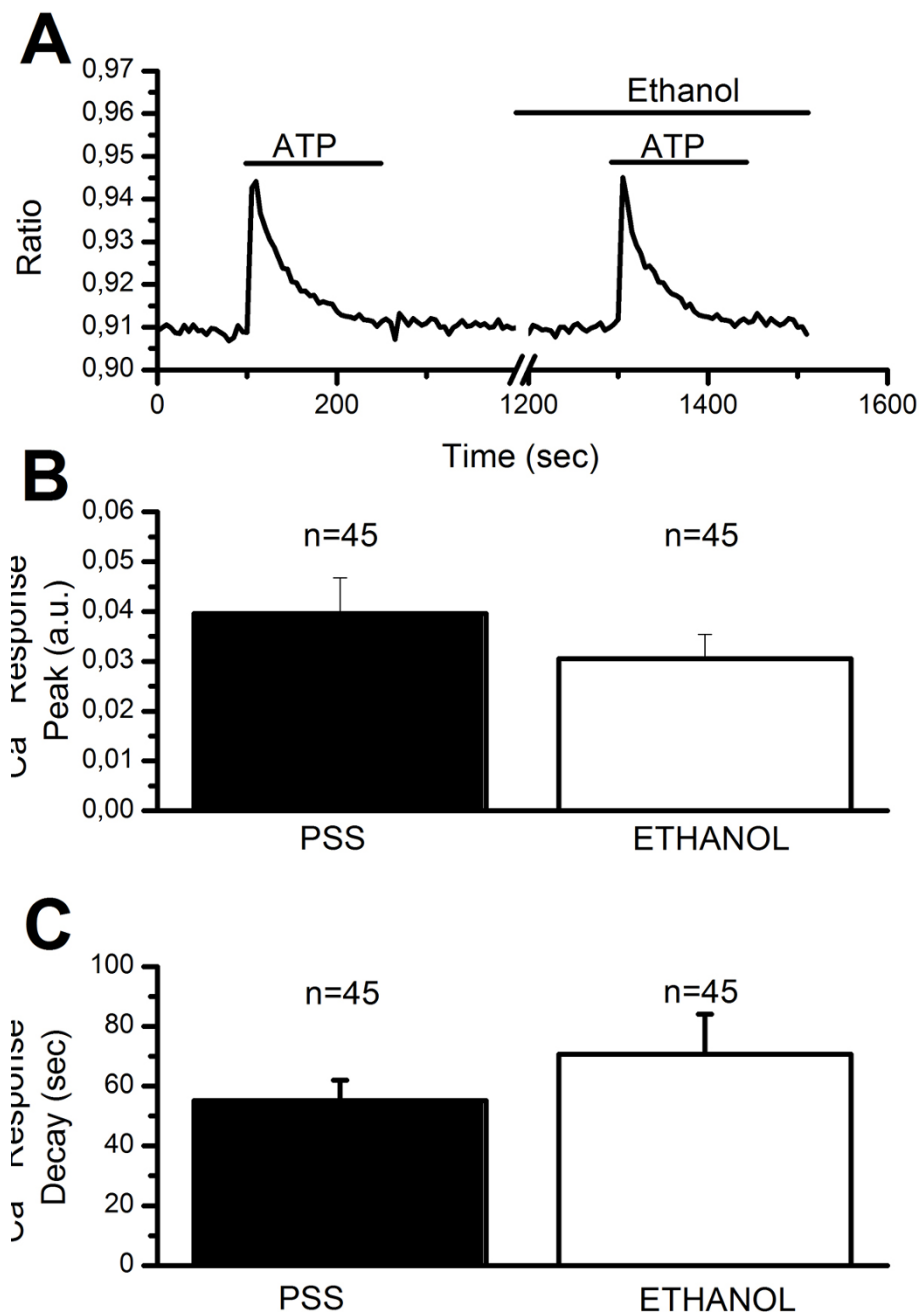


Figure 4 Ethanol does not significantly affect the Ca²⁺ response to ATP. A, 20 min pre-incubation with octanol (4 mM) does not remarkably affect either the amplitude or the kinetics of the Ca²⁺ response to ATP (20 μM). The trace is the average of 16 cells from the same microscopic field of view. B, mean±SE of the peak of ATP-induced elevation in [Ca²⁺]. The asterisk indicates a level of significant difference <0.05. The number of ECs analyzed in each condition is indicated above the bars. C, B, mean±SE of the rate of decay of ATP-induced Ca²⁺ signal. The decay was measured by fitting the Ca²⁺ tracings with a first order exponential equation (OriginPro 7, OriginLab Corp, Northampton MA). The asterisk indicates a level of significant difference <0.05. The number of ECs analyzed in each condition is indicated above the bars.

suppression of specific Cx isoforms is not feasible, such as in rat endothelium [33,45]. In addition, Cx-mimetic peptides do not present the drawbacks reported in Cx-deficient mice [53]. For instance, Cx40^{-/-} mice manifest either higher [54] or lower [55] levels of Cx37 as well as a

significant decrease in Cx43 expression [56]. This feature makes the interpretation of the physiological data collected from these animals difficult. Incubating the aortic rings for 1 hour with either ^{37,43}Gap27 (300 μM) or ⁴⁰Gap27 (300 μM) significantly (p < 0.05) reduced injury-

evoked NOP (Figure 5A and Figure 5B). The statistical analysis of these experiments has been reported in Figure 5C and Figure 5D. NO signalling was unaffected by peptides corresponding to scrambled sequences in Gap27, indicating that the inhibition resulted from specific sequence recognition on the selected Cx isotypes (Figure 5C and Figure 5D). Therefore, it is reasonable to conclude that Ca^{2+} entry driving eNOS activity and NO liberation at the wound edge is supported by Cx37Hcs, Cx40Hcs, and Cx43Hcs. As discussed elsewhere [57], eNOS activation requires a sub-plasmalemmal Ca^{2+} increase rather a bulk increase in cytosolic Ca^{2+} levels. Such a localized Ca^{2+} event may not even be detected by conventional epifluorescence microscopy [58]. As a consequence, we went on in assessing whether Cx-mimetic peptides targeting Cx37, Cx40, and Cx43 were also able to reduce the long-lasting decay phase of the Ca^{2+}

response to mechanical damage. Figures 6A to 6D depict that neither $^{37,43}Gap27$ (300 μM) nor $^{40}Gap27$ (300 μM) significantly reduce either the initial Ca^{2+} peak or the subsequent decay phase. Rather, both Cx-mimetic peptides increased the amplitude of the initial Ca^{2+} response (Figure 6A-6C). Consistently, acute application of both peptides during recovery of the Ca^{2+} signal towards the baseline did not decrease the $[Ca^{2+}]_i$ ($n = 31$, $^{40}Gap27$; $n = 52$, $^{37,43}Gap27$) (not shown). The same results were found by extending the pre-incubation period with both $^{37,43}Gap27$ and $^{40}Gap27$ up to 2 hours (Figure 6B-6D) [49]. Taken as a whole, these results suggest that Cx37Hcs, Cx40Hcs, and Cx43Hcs mediate the sub-membranal Ca^{2+} elevation responsible for eNOS activation, but not the bulk increase in $[Ca^{2+}]_i$ caused by mechanical damage in surviving ECs. As the pharmacological profile of the decay phase is consistent with a CxHc

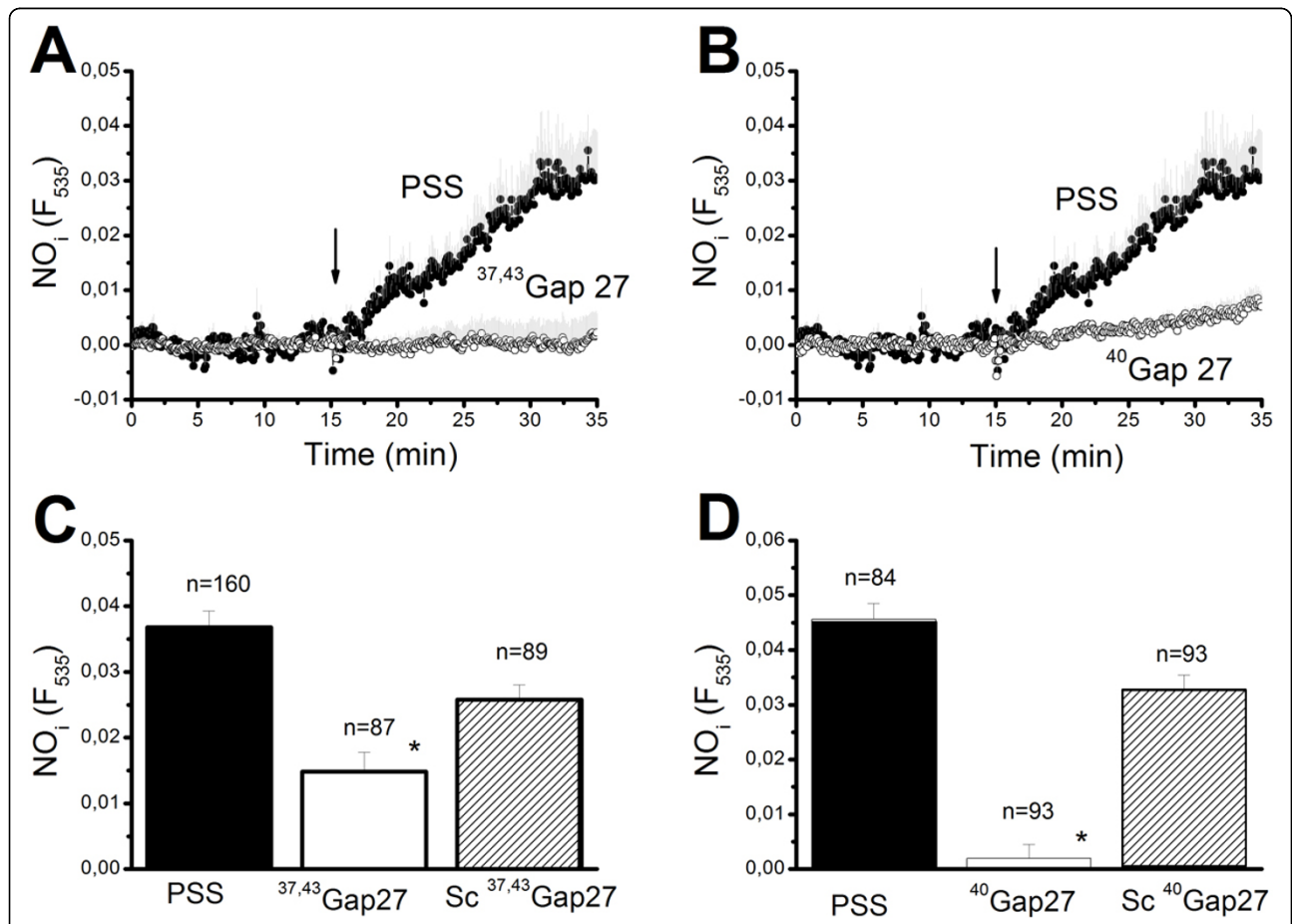
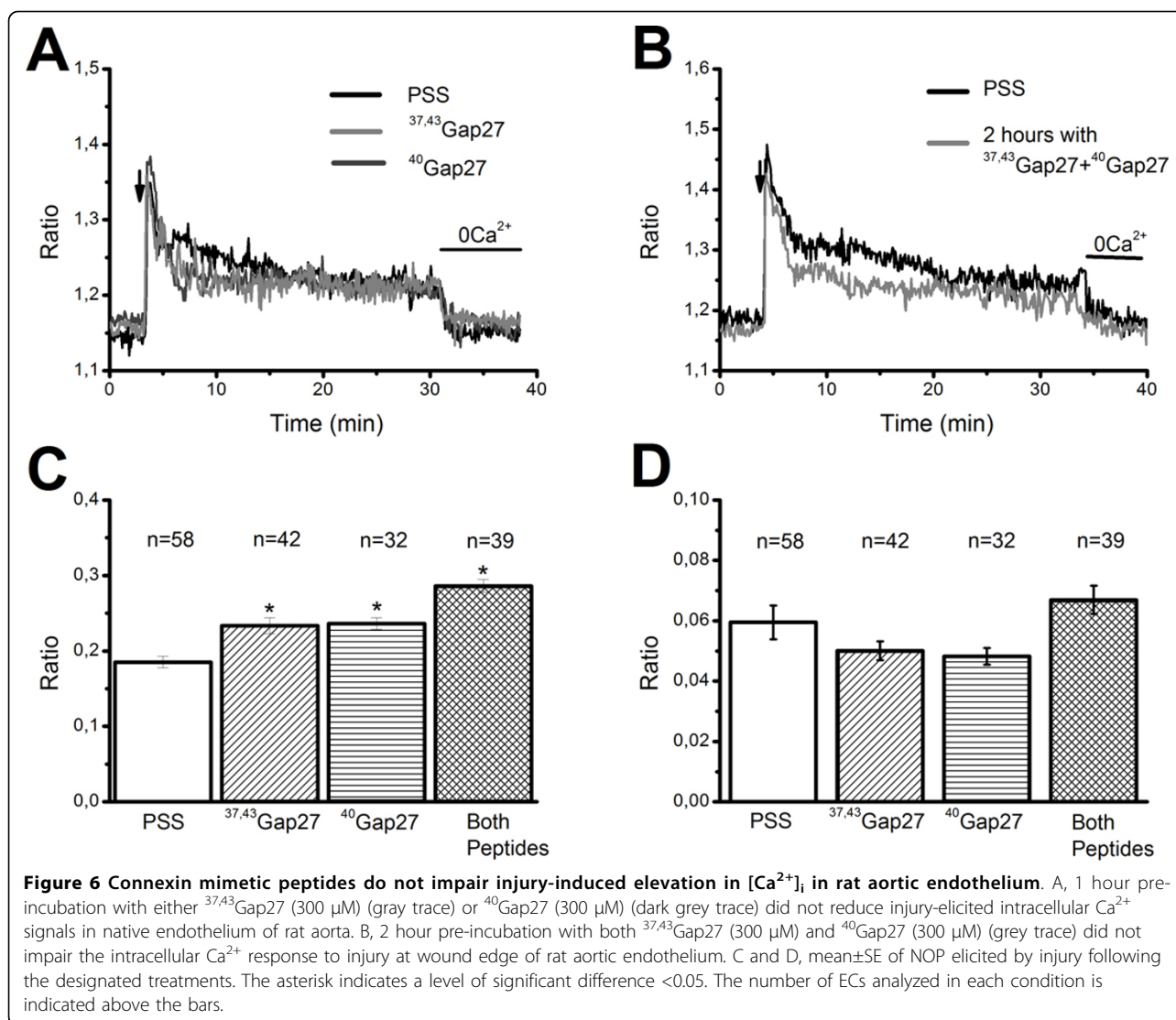


Figure 5 Connexin-mimetic peptides affect injury-induced NO synthesis. 1 hour pre-incubation with either $^{37,43}Gap27$ (300 μM) (A) or $^{40}Gap27$ (300 μM) (B) dramatically attenuated the increase in intracellular NO levels provoked by endothelial lesion. For sake of clarity, the control curve has been depicted in each Panel. All the experiments depicted in this figure have been carried out on the same day on different rings isolated from the same animal. The control (PSS, closed circles) trace is the mean \pm SE of 9 cells, while the $^{37,43}Gap27$ (open circles in Panel A) and $^{40}Gap27$ traces (open circles in Panel B) are, respectively, the mean \pm SE of 5 and 7 cells. C and D, mean \pm SE of NOP elicited by injury following the designated treatments. Each drug was applied was 20 min before carrying out the lesion. The asterisk indicates a level of significant difference <0.05 . The number of ECs analyzed in each condition is indicated above the bars. See the text for drug concentrations



pathway [2,26,27], it is conceivable that one or more additional Cx isoforms are expressed in rat aortic endothelium.

NO synthesis is abolished by caveolar disruption with M β CD

NO production by a highly localized Ca^{2+} signal sneaking beneath the plasma membrane requires the spatial proximity between the Ca^{2+} source and eNOS [57,58]. Caveolae represent a signalling platform responsible for translating Ca^{2+} entry into specific signal transduction pathways [24,57]. M β CD is a cholesterol-binding agent which has been widely employed to disrupt caveolar integrity in a variety of cell types, including ECs [24]. We first probed M β CD (10 mM) efficacy in impairing caveolae-dependent signalling by testing its effect on SOCE. As shown elsewhere [27,35], SOCE sustains the

Ca^{2+} response to high concentrations of ATP (300 μM) in the endothelium of excised rat aorta. As shown in Figure 7A, M β CD significantly reduced the amplitudes of both the initial Ca^{2+} peak and the subsequent plateau, which are both supported by SOCE [27]. Consistently, M β CD inhibited ATP-dependent NOP in 85 out of 85 cells, whereas 93 out of 93 untreated cells liberated NO when exposed to ATP (Figure 7B). Then, to assess the role of caveolae in coupling CxHcs-mediated Ca^{2+} inflow with eNOS activation, rat aortic rings were pre-incubated (30 min) in the presence of M β CD (10 mM). Exposition to M β CD did not significantly affect the Ca^{2+} response to injury (Figure 8A and Figure 8B), but prevented the downstream NO synthesis (Figure 8C and Figure 8D). Collectively, these data indicate that the proximity between CxHcs and eNOS is maintained by the caveolar

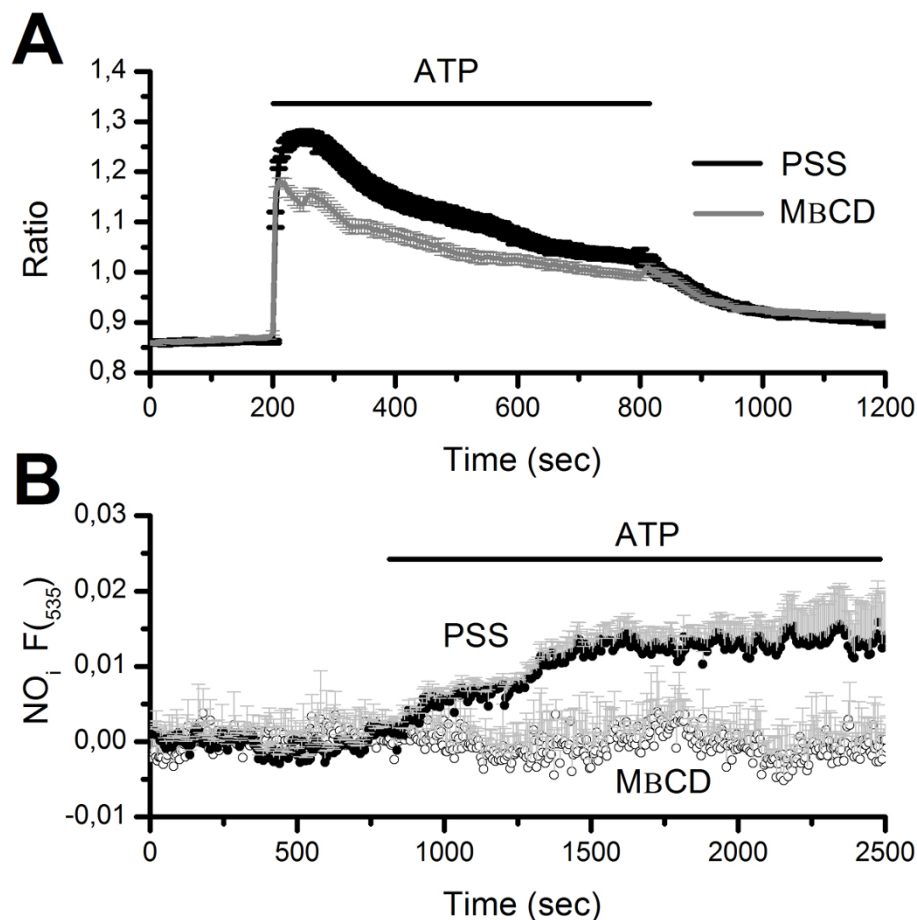


Figure 7 Caveolar impairment impairs both the Ca²⁺ response to ATP and the accompanying NO production in rat aortic endothelium. A, pre-treatment with 10 mM MβCD to disrupt caveolae significantly hinders both the magnitude and the plateau of the Ca²⁺ response to ATP (300 μM). Both phases depend on store-dependent Ca²⁺ inflow when ATP is administered at this dose. The traces are the mean ±SE of 220 (PSS, black trace) and 186 cells (MβCD, red trace), respectively, from no less than three different animals. B, MβCD (10 mM) suppresses ATP-induced NO synthesis. The traces are the mean ±SE, respectively, of 150 (PSS, closed circles) and 4 (MβCD, open circles) cells from no less than three different animals.

signalling platform within the plasma membrane. This feature is consistent with the highly localized nature of the Ca²⁺ signal driving NO synthesis.

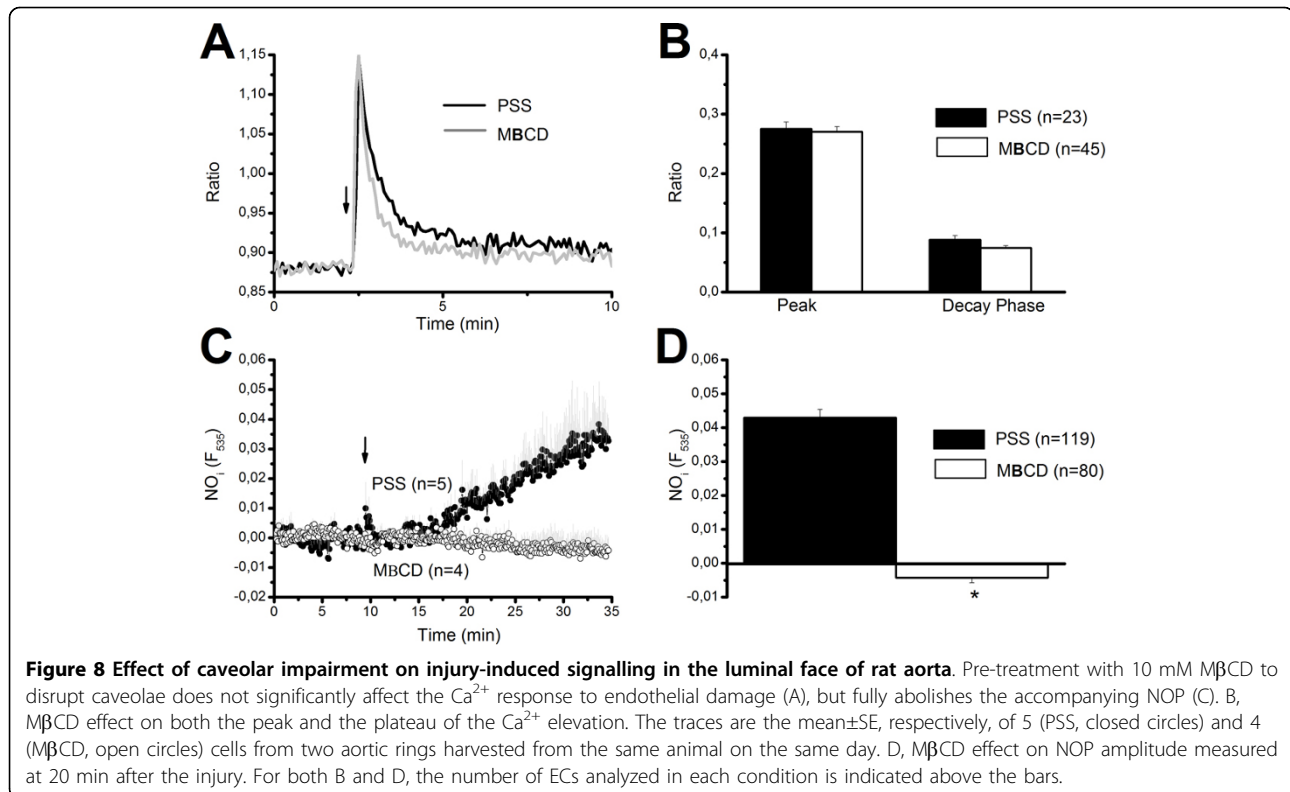
Discussion

Endothelial-derived NO drives a number of processes which protects the vessel wall by pro-atherosclerotic events and maintain vascular homeostasis. NO exerts both an anti-aggregating and anti-inflammatory action, promotes endothelial survival, proliferation and migration, and prevents the phenotypic remodeling of the underlying layer of SMCs [2,19,59]. The present investigation provides the first evidence that endothelial ablation similar to that induced *in vivo* by stent deployment causes a local increase in NO levels by activating the ECs immediately adjacent to the lesion site. We further elucidated the molecular machinery responsible for

eNOS recruitment and showed that injury-induced NOP is triggered by Ca²⁺ entry through Cx37Hcs, Cx40Hcs, and Cx43Hcs. The coupling between Ca²⁺ inflow and eNOS activation might be provided by cholesterol enriched membrane microdomains, such as caveolae. These data shed novel light on the endothelial reaction to mechanical scraping and hint at the Ca²⁺ machinery as a novel target to restore endothelial integrity after reconstructive surgery and to treat vascular diseases [2,12].

NO synthesis in the injured endothelium of rat aorta is supported by CxHcs-mediated Ca²⁺ entry

The procedure that we adopted for mechanical scraping mimics the partial denudation of the endothelial monolayer that has been described upon stent deployment in a number of pre-clinical models [41-43]. The extent of the lesion we inferred is smaller as compared to the degree



of endothelial ablation observed after coronary stenting in humans in patients [60], albeit it is in the same range as that described in previous studies addressing the role of Ca²⁺ signals in the regeneration of wounded monolayers [2,12]. However, a focal region devoid of the EC covering is rapidly repaired by neighbouring ECs, which spread into the lesioned area and undergo mitosis [61]. Accordingly, the daily rate of EC replication is increased by 50% at these sites in the rat aorta [62]. Therefore, the approach we utilized to perform the endothelial injury is useful to: 1) get insights into the acute response to the arterial damage in terms of Ca²⁺ response and accompanying NO synthesis and 2) elucidate the molecular machinery driving the subsequent process of endothelial regeneration. Three pieces of evidences suggest that injury-induced NO synthesis is mainly driven by Ca²⁺ influx. First, there is no detectable NOP when the endothelium is scraped in absence of extracellular Ca²⁺. Second, injury-elicited NOP is not significantly affected by depletion of the InsP₃-sensitive Ca²⁺ stores with thapsigargin, although we cannot rule out the possibility that InsP₃-sensitive NOP falls below the resolution of our fluorimetric system. Third, blockade of Ca²⁺ entry by classic gap junction blockers, but not by BTP-2, dampen NO synthesis. These observations are consistent with the Fura-2 data and strongly suggest that CxHcs, rather than SOCE, provide the Ca²⁺ source driving NO liberation in

damaged endothelium. Rat aortic endothelium has been found to express three main Cx isoforms, namely Cx37, Cx40, and Cx43 [32,49]. In order to elucidate which of them underpins NO synthesis, we exploited the reported selectivity of Cx-mimetic peptides [33,45]. Both ^{37,43}Gap27 and ⁴⁰Gap27 significantly attenuated NOP triggered by Ca²⁺ entry at the lesion site. The following lines of evidence confirm the specificity of ^{37,43}Gap27 and ⁴⁰Gap27: 1) pre-treatment with scrambled peptides did not detectably alter injury-induced NO synthesis (present study); and 2) ^{37,43}Gap27 and ⁴⁰Gap27 do not alter endogenous expression of Cx40 and Cx43 in the smooth muscle cell line A7r5, as well as connexin trafficking and *de novo* formation of gap plaques in A7r5 cells expressing Cx43-GFP [49]. These data strongly suggest that injury-elicited NOP at the wound edge of the aortic surface is sustained by Ca²⁺ influx through Cx37Hcs, Cx40Hcs, and Cx43Hcs. These results are further supported by previous studies, which showed that Cx43-containing gap junctions mediate the spreading of pro-inflammatory Ca²⁺ waves in lung capillaries [36] and sustain ATP-promoted Ca²⁺ bursts in uterine artery ECs [63]. Consistent with the role served by CxHc as Ca²⁺ entry pathways in vascular endothelium, Cx37Hc and Cx43Hc were recently shown to maintain bradykinin-induced intracellular Ca²⁺ oscillations in rat brain endothelial cells [29], while Cx32Hc- and Cx43Hc-mediated Ca²⁺ inflow supports the spiking response to

bradykinin in Madin-Darby canine kidney cells [30]. Similarly, Cx43Hcs has already been shown to mediate Ca^{2+} influx and activate Ca^{2+} -dependent downstream targets in a variety of cell types other than endothelial cells [31,64].

Caveolar integrity is required to support injury-dependent NO synthesis in rat aortic endothelium

The subcellular analysis of the Ca^{2+} signal driving eNOS activation has shown that Ca^{2+} entry results in a highly localized Ca^{2+} microdomain which is restricted to the inner leaflet of the plasma membrane [57,58]. Therefore, CxHcs and eNOS are likely to tightly interact in order for Ca^{2+} inflow to stimulate NOP in injured endothelium. This hypothesis is supported by a number of recent studies. First, eNOS physically interacts with both Cx37 and Cx40 in *in situ* ECs of mouse aorta [65]. Second, Cx43 colocalizes with eNOS in native ECs covering the lumen of mice thoracodorsal arteries [66]. Third, Cx37 coimmunoprecipitates with eNOS in a number of human primary ECs [67]. The physical coupling between CxHcs and eNOS might be provided by cholesterol enriched membrane domains, such as caveolae. Indeed, M β CD abrogated injury-triggered NOP without affecting the underpinning Ca^{2+} signal. This feature suggests that disruption of caveolae does not modify CxHcs gating, but physically displaces eNOS from the local source of Ca^{2+} responsible for its engagement. Consistently, caveolin-1 regulates the correct trafficking and localization to the plasma membrane of several Cx isoforms, including Cx 37, 40, and 43 [66,68,69]. Moreover, the expression of Cx37, Cx40 and Cx43 is decreased in caveolin-1 knock out arteries as compared to wild-type mice [70]. On the other hand, eNOS is targeted to caveolae by cotranslational N-myristoylation and posttranslational palmitoylation and interacts with the scaffolding domain of caveolin-1 [23]. It is, therefore, likely that caveolae maintain CxHcs in close proximity to eNOS and, perhaps, to other relevant signalling proteins, as recently suggested in [70]. Interestingly, while ineffective in regulating the Ca^{2+} -mobilizing properties of CxHcs, caveolar integrity is key to SOCE recruitment in vascular ECs [57]. In line with these evidences, cholesterol depletion with M β CD prevents store-operated Ca^{2+} inflow and NOP triggered by ATP in native rat aortic endothelium.

The putative role of NO in lesioned endothelium

The increase in NO levels at the wound edge represents an immediate, localized self-protecting response of the vessel wall to EC loss. Indeed, the NO signal is confined within the area experiencing the elevation in $[\text{Ca}^{2+}]_i$ induced by the injury and does not spread farther away (see Figure 2C). EC detachment due to physiological turnover or pathological events causes the exposition of patches of sub-endothelial matrix which may activate platelets, a process known to promote an inflammatory reaction which favours the onset of atherosclerosis [71].

However, increased local NO synthesis inhibits both platelet aggregation and VSMC proliferation and migration, thus adversely affecting both neointima formation and inflammation [1,19]. NO might play a key role also in subsequent healing [19,59]. Accordingly, NO inhibits apoptosis and stimulates proliferation and migration, which are an essential pre-requisite for ECs to spread and cover the de-endothelized area [19,59]. In injured rat aortic rings, both eNOS protein and enzyme activity have been shown to augment at the proliferating wound edge, but not in ECs distant from the lesion [25]. Moreover, aortic segments obtained from eNOS knock-out (KO) mice and cultured in Matrigel displayed a dramatic reduction in EC sprouting and proliferation as compared to wild-type animals [72]. Consistent with these *in vitro* observations, intravenous infusion of a NO donor accelerated *in vivo* regeneration of the endothelium lining the lumen of injured rat carotid artery [73]. In addition, neo-vascularization of ischemic hindlimbs, which depends on EC proliferation and migration, is significantly reduced in eNOS KO mice [74] and enhanced in eNOS overexpressing mice [75]. Finally, NO liberated by cells at the wound edge might recruit circulating endothelial progenitor cells and speed up the healing process [76].

The changes in DAF-FM fluorescence measured from the endothelial layer could also reflect some NOP occurring within the underlying SMCs, which generate a Ca^{2+} wave in response to mechanical injury [48]. This Ca^{2+} signal might, in turn, activate SMC eNOS and contribute to the increase in DAF-FM fluorescence we recorded from ECs at the wound edge [77]. However, injury-induced Ca^{2+} waves in SMCs entirely depend on intracellular Ca^{2+} release, rather than Ca^{2+} influx [48]. As a consequence, the fact that all the protocols impairing injury-elicited Ca^{2+} inflow into ECs nearby the lesion site (i.e. removal of external Ca^{2+} and exposition to gap junction blockers) affected the accompanying increase in DAF-FM fluorescence strongly hint at the endothelial origin of the NO signal. Nevertheless, the possibility that NO synthesis also takes place within the tunica media of damaged rat aortic rings cannot be ruled out.

Evidence for the expression of additional Cx isoforms in the endothelium of rat aorta

The results presented in the current investigation support a role for Cx37Hcs, Cx40Hcs, and Cx43Hcs in driving the Ca^{2+} -dependent eNOS activation at the wound edge of rat aortic endothelium. However, the decay phase of the Ca^{2+} response to injury is not significantly affected by 1 hour pre-treatment with Cx-mimetic peptides. This feature concurs with the highly localized, sub-membranal nature of the Ca^{2+} signal engaging eNOS in vascular ECs [57,58], which are

detected by genetically-encoded Ca^{2+} fluorophores selectively targeting the caveolae, but not the bulk cytoplasm [58]. This finding raises us to wonder about the molecular nature of the membrane pathway(s) responsible for the massive Ca^{2+} influx observed during the decay phase. As widely discussed elsewhere [2,12], the pharmacological profile of such a route is typical of a CxHc. The lack of effect of Cx-mimetic peptides on the bulk increase in $[\text{Ca}^{2+}]_i$ that features the decay phase of the Ca^{2+} response to injury, therefore, suggests that Cx isoforms other than Cx 37, 40, and 43 are expressed in the native endothelium of rat aorta. This hypothesis is strongly supported by the recent discovery of Cx32 mRNA and protein in both cultured and blood vessel ECs [50]. Cx32Hcs, in particular, has recently been shown to mediate Ca^{2+} influx when transfected in HeLa cells [78] and in Madin-Darby canine kidney cells [30]. Additional Cx isoforms that have been implicated in Ca^{2+} entry across the plasmalemma are Cx26 [79] and Cx45 [80]. That Cx isoforms other than Cx37, Cx40, and Cx43 are expressed in the vessel wall has also been proposed by Tang and Vanhoutte [52] and by Sorensen et al. [81]. Both groups reached the same conclusion upon the finding that connexin mimetic peptides failed to reproduce the effects of classic gap junction blockers in the vasculature. Recent evidence hinted at pannexins (Pxs), which are hexameric channels structurally unrelated to Cxs, as the molecular substrates for the hemichannels (the so-called pannexons) present in the non-junctional area of cell membrane [82]. However, to the best of our knowledge, Px expression has hitherto been reported only in microvascular beds rather than in macrovessels, such as aorta [82]. Moreover, as recently pointed out [2,12], Px hemichannels are unaffected by La^{3+} and heptanol, which block CxHcs and impair injury-induced Ca^{2+} entry in the intact endothelium of rat aorta [26,27]. Accordingly, our preliminary data indicate that probenecid, a widely employed Px blocker, does not significantly affect the Ca^{2+} response to mechanical injury in rat aortic endothelium. It is worth of noting that Cx-mimetic peptides, either separately or in combination, caused an increase in the InsP_3 -dependent initial Ca^{2+} peak. A recent study reported that CxHcs may mediate InsP_3 efflux into the extracellular space [83]. It is, therefore, conceivable that the prolonged pre-treatment with Cx-mimetic peptides prevented InsP_3 produced at the wound edge from moving out of the cytosol, thus increasing the extent of Ca^{2+} release in the first raw of cells.

Conclusion

In conclusion, this study provides the first clear-cut evidence that arterial damage activates surviving ECs to produce NO by inducing Ca^{2+} entry via Cx37Hcs,

Cx40Hcs, and Cx43Hcs. Our data indicate that the membranal compartmentalization of CxHcs and eNOS depends on is enabled by caveolae. Experiments are under way in our laboratory to assess the role played by CxHcs-induced NO synthesis in the regeneration of rat aortic endothelium. These results outline the importance of designing novel prosthetic devices able to release NO after vascular interventions, such as arterial bypass grafting and intravascular stent deployment [84]. Indeed, the currently employed drug-eluting stents may result in late arterial occlusion and, eventually, in patient death due to their reported capability to inhibit endothelial regeneration [11]. Alternatively, the signaling cascade leading to NO production that we have described in the present work might be the alternative target of novel drug-eluting stents. The cardiological practice has unveiled a variety of compounds, such as ZP123 or rotigaptide and (2S,4R)-1-(2-aminoacetyl)-4-benzamidopyrrolidine-2-carboxylic acid hydrochloride (GAP-134), which increase CxHc conductance and, therefore, might be exploited to increase Ca^{2+} influx at the lesion site [2,12]. These substances have successfully been probed as orally-active and anti-arrhythmic drugs in Phase I and II clinical trials conducted on patients suffering either from atrial fibrillation or from unstable angina or myocardial infarction [85,86]. In perspective, these molecules might be locally released by drug-eluting stents to improve CxHc-mediated signalling and boost NO synthesis after surgical interventions on diseased vessels [2,12].

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

RBR: conceived the study, carried out the experiments, analyzed and interpreted the data, drafted the manuscript. JEAC: conceived the study, carried out the experiments, analyzed and interpreted the data. AR: conceived the study, carried out the experiments, analyzed and interpreted the data. ADC: critically revised the manuscript. MPC: critically revised the manuscript. SM: critically revised the manuscript. GG: conceived the study and critically revised the manuscript. FM: conceived the study, analyzed and interpreted the data and drafted the manuscript. FT: conceived the study, analyzed and interpreted the data, critically revised the drafted manuscript. All authors read and approved the final manuscript.

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Declarations

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References

1. Tesfamariam B, DeFelice AF: Endothelial injury in the initiation and progression of vascular disorders. *Vascular Pharmacology* 2007, **46**(4):229-237.
2. Moccia F, Billington RA, Santella L: Pharmacological characterization of NAADP-induced Ca²⁺ signals in starfish oocytes. *Biochemical and Biophysical Research Communications* 2006, **348**:329-336.
3. Vanhoutte PM: Endothelial Dysfunction - The First Step Toward Coronary Arteriosclerosis. *Circulation Journal* 2009, **73**(4).
4. Yoder M, Rounds S: Bad blood, bad endothelium: ill fate? *Blood* 2011, **117**(13):3479-3480.
5. Dragoni S, Laforenza U, Bonetti E, Lodola F, Bottino C, Berra-Romani R, Bongio GC, Cinelli MP, Guerra G, Pedrazzoli P, et al: Vascular Endothelial Growth Factor Stimulates Endothelial Colony Forming Cells Proliferation and Tubulogenesis by Inducing Oscillations in Intracellular Ca²⁺ Concentration. *Stem Cells* 2011, **29**(11):1898-1907.
6. Sanchez-Hernandez Y, Laforenza U, Bonetti E, Fontana J, Dragoni S, Russo M, Avelino-Cruz JE, Schinelli S, Testa D, Guerra G, et al: Store-Operated Ca²⁺ Entry Is Expressed in Human Endothelial Progenitor Cells. *Stem Cells and Development* 2010, **19**(12):1967-1981.
7. Lodola F, Laforenza U, Bonetti E, Lim D, Dragoni S, Bottino C, Ong HL, Guerra G, Ganini C, Massa M, et al: Store-operated Ca²⁺ entry is remodelled and controls in vitro angiogenesis in endothelial progenitor cells isolated from tumoral patients. *PLoS one* 2012, **7**(9):e42541.
8. Moccia F, Dragoni S, Lodola F, Bonetti E, Bottino C, Guerra G, Laforenza U, Rosti V, Tanzi F: Store-Dependent Ca²⁺ Entry in Endothelial Progenitor Cells As a Perspective Tool to Enhance Cell-Based Therapy and Adverse Tumour Vascularization. *Current Medicinal Chemistry* 2012, **19**(34):2561-2580.
9. Piscopo S, Moccia F, Di Cristo C, Caputi L, Di Cosmo A, Brown ER: Pre- and postsynaptic excitation and inhibition at octopus optic lobe photoreceptor terminals; implications for the function of the 'presynaptic bags'. *European Journal of Neuroscience* 2007, **26**:2196-2203.
10. Moccia F, Bonetti E, Dragoni S, Fontana J, Lodola F, Berra Romani R, Laforenza U, Rosti V, Tanzi F: Hematopoietic Progenitor and Stem Cells Circulate by Surfing on Intracellular Ca²⁺ Waves: A Novel Target for Cell-based Therapy and Anti-cancer Treatment? *Current Signal Transduction Therapy* 2012, **7**(2).
11. Zargham R: Preventing restenosis after angioplasty: a multistage approach. *Clinical Science* 2008, **114**(3-4):257-264.
12. Moccia F, Di Cristo C, Winlow W, Di Cosmo A: GABA(A)- and AMPA-like receptors modulate the activity of an identified neuron within the central pattern generator of the pond snail *Lymnaea stagnalis*. *Invertebrate Neuroscience* 2009, **9**(1):29-41.
13. Siddique A, Shantsila E, Lip GH, Varma C: Endothelial progenitor cells: what use for the cardiologist? *Journal of angiogenesis research* 2010, **2**:6-6.
14. Finn AV, Nakazawa G, Joner M, Kolodgie FD, Mont EK, Gold HK, Virmani R: Vascular responses to drug eluting stents - Importance of delayed healing. *Arteriosclerosis Thrombosis and Vascular Biology* 2007, **27**(7).
15. Otsuka F, Finn AV, Yazdani SK, Nakano M, Kolodgie FD, Virmani R: The importance of the endothelium in atherothrombosis and coronary stenting. *Nature Reviews Cardiology* 2012, **9**(8).
16. Testa D, Guerra G, Marcuccio G, Landolfo PG, Motta G: Oxidative stress in chronic otitis media with effusion. *Acta oto-laryngologica* 2012, **132**(8):834-837.
17. Cattaneo F, Iaccio A, Guerra G, Montagnani S, Ammendola R: NADPH-oxidase-dependent reactive oxygen species mediate EGFR transactivation by FPRL1 in WKY/Mv-stimulated human lung cancer cells. *Free radical biology and medicine* 2011, **51**(6):1126-1136.
18. Conti V, Russomanno G, Corbi G, Guerra G, Grasso C, Filippelli W, Paribello V, Ferrara N, Filippelli A: Aerobic training workload affects human endothelial cells redox homeostasis. *Medicine and science in sports and exercise* 2013, **45**(4):644-653.
19. Mancardi D, Pla AF, Moccia F, Tanzi F, Munaron L: Old and New Gasotransmitters in the Cardiovascular System: Focus on the Role of Nitric Oxide and Hydrogen Sulfide in Endothelial Cells and Cardiomyocytes. *Current Pharmaceutical Biotechnology* 2011, **12**(9):1406-1415.
20. Al-Sa'doni HH, Ferro A: S-nitrosothiols as nitric oxide-donors: Chemistry, biology and possible future therapeutic applications. *Current Medicinal Chemistry* 2004, **11**(20):2679-2690.
21. Yao XQ, Huang Y: From nitric oxide to endothelial cytosolic Ca²⁺: a negative feedback control. *Trends in Pharmacological Sciences* 2003, **24**(6):263-266.
22. Moccia F, Berra-Romani R, Tanzi F: Update on vascular endothelial Ca²⁺ signalling: A tale of ion channels, pumps and transporters. *World journal of biological chemistry* 2012, **3**(7).
23. Sbaa E, Frerart F, Feron O: The double regulation of endothelial nitric oxide synthase by caveolae and caveolin: A paradox solved through the study of angiogenesis. *Trends in Cardiovascular Medicine* 2005, **15**(5):157-162.
24. Xu Y, Bulikema H, van Gilst WH, Henning RH: Caveolae and endothelial dysfunction: Filling the caves in cardiovascular disease. *European Journal of Pharmacology* 2008, **585**(2-3):256-260.
25. Poppa V, Miyashiro JK, Corson MA, Berk BC: Endothelial NO synthase is increased in regenerating endothelium after denuding injury of the rat aorta. *Arteriosclerosis Thrombosis and Vascular Biology* 1998, **18**(8):1312-1321.
26. Berra-Romani R, Raqeeb A, Torres-Jacome J, Guzman-Silva A, Guerra G, Tanzi F, Moccia F: The Mechanism of Injury-Induced Intracellular Calcium Concentration Oscillations in the Endothelium of Excised Rat Aorta. *Journal of Vascular Research* 2012, **49**(1):65-76.
27. Berra-Romani R, Raqeeb A, Avelino-Cruz JE, Moccia F, Oldani A, Speroni F, Taglietti V, Tanzi F: Ca²⁺ signaling in injured in situ endothelium of rat aorta. *Cell Calcium* 2008, **44**(3):298-309.
28. Verma V, Hallett MB, Leybaert L, Martin PE, Evans WH: Perturbing plasma membrane hemichannels attenuates calcium signalling in cardiac cells and HeLa cells expressing connexins. *European Journal of Cell Biology* 2009, **88**(2).
29. De Bock M, Culot M, Wang N, Bol M, Decrock E, De Vuyst E, da Costa A, Dauwe I, Vinken M, Simon AM, et al: Connexin channels provide a target to manipulate brain endothelial calcium dynamics and blood-brain barrier permeability. *Journal of Cerebral Blood Flow and Metabolism* 2011, **31**(9).
30. De Bock M, Wang N, Bol M, Decrock E, Ponsaerts R, Bultynck G, Dupont G, Leybaert L: Connexin 43 Hemichannels Contribute to Cytoplasmic Ca²⁺ Oscillations by Providing a Bimodal Ca²⁺-dependent Ca²⁺ Entry Pathway. *Journal of Biological Chemistry* 2012, **287**(15).
31. Park JH, Lee MY, Heo JS, Han HJ: A potential role of connexin 43 in epidermal growth factor-induced proliferation of mouse embryonic stem cells: Involvement of Ca²⁺/PKC, p44/42 and p38 MAPKs pathways. *Cell Proliferation* 2008, **41**(5):786-802.
32. Gabriels JE, Paul DL: Connexin43 is highly localized to sites of disturbed flow in rat aortic endothelium but connexin37 and connexin40 are more uniformly distributed. *Circulation Research* 1998, **83**(6):636-643.
33. Griffith TM: Endothelium-dependent smooth muscle hyperpolarization: do gap junctions provide a unifying hypothesis? *British Journal of Pharmacology* 2004, **141**(6):881-903.

34. Berra-Romani R, Rinaldi C, Raqeeb A, Castelli L, Magistretti J, Taglietti V, Tanzi F: **The duration and amplitude of the plateau phase of ATP- and ADP-evoked Ca²⁺ signals are modulated by ectonucleotidases in situ endothelial cells of rat aorta.** *Journal of Vascular Research* 2004, **41**(2):166-173.
35. Berra-Romani R, Raqeeb A, Guzman-Silva A, Torres-Jacome J, Tanzi F, Moccia F: **Na⁺-Ca²⁺ exchanger contributes to Ca²⁺ extrusion in ATP-stimulated endothelium of intact rat aorta.** *Biochemical and Biophysical Research Communications* 2010, **395**(1):126-130.
36. Moccia F, Bertoni G, Pla AF, Dragoni S, Pupo E, Merlino A, Mancardi D, Munaron L, Tanzi F: **Hydrogen Sulfide Regulates Intracellular Ca²⁺ Concentration in Endothelial Cells From Excised Rat Aorta.** *Current Pharmaceutical Biotechnology* 2011, **12**(9):1416-1426.
37. Pupo E, Pla AF, Avanzato D, Moccia F, Cruz J-EA, Tanzi F, Merlino A, Mancardi D, Munaron L: **Hydrogen sulfide promotes calcium signals and migration in tumor-derived endothelial cells.** *Free Radical Biology and Medicine* 2011, **51**(9):1765-1773.
38. Koyama T, Kimura C, Park SJ, Oike M, Ito Y: **Functional implications of Ca²⁺ + mobilizing properties for nitric oxide production in aortic endothelium.** *Life Sciences* 2002, **72**(4-5):511-520.
39. Hartell NA, Archer HE, Bailey CJ: **Insulin-stimulated endothelial nitric oxide release is calcium independent and mediated via protein kinase B.** *Biochemical Pharmacology* 2005, **69**(5):781-790.
40. Dedkova EN, Blatter LA: **Nitric oxide inhibits capacitative Ca²⁺ entry and enhances endoplasmic reticulum Ca²⁺ uptake in bovine vascular endothelial cells.** *Journal of Physiology-London* 2002, **539**(1):77-91.
41. Rogers C, Parikh S, Seifert P, Edelman ER: **Endogenous cell seeding - Remnant endothelium after stenting enhances vascular repair.** *Circulation* 1996, **94**(11):2909-2914.
42. Orford JL, Selwyn AP, Ganz P, Popma JJ, Rogers C: **The comparative pathobiology of atherosclerosis and restenosis.** *American Journal of Cardiology* 2000, **86**(4B).
43. Rogers C, Tseng DY, Squire JC, Edelman ER: **Balloon-artery interactions during stent placement - A finite element analysis approach to pressure, compliance, and stent design as contributors to vascular injury.** *Circulation Research* 1999, **84**(4):378-383.
44. Schalper KA, Palacios-Prado N, Orellana JA, Saez JC: **Currently used methods for identification and characterization of hemichannels.** *Cell Communication and Adhesion* 2008, **15**(1-2):207-218.
45. Evans WH, De Vuyst E, Leybaert L: **The gap junction cellular internet: connexin hemichannels enter the signalling limelight.** *Biochemical Journal* 2006, **397**:1-14.
46. Moccia F, Berra-Romani R, Baruffi S, Spaggiari S, Signorelli S, Castelli L, Magistretti J, Taglietti V, Tanzi F: **Ca²⁺ uptake by the endoplasmic reticulum Ca²⁺-ATPase in rat microvascular endothelial cells.** *Biochemical Journal* 2002, **364**:235-244.
47. Christ GJ, Spektor M, Brink PR, Barr L: **Further evidence for the selective disruption of intercellular communication by heptanol.** *American Journal of Physiology-Heart and Circulatory Physiology* 1999, **276**(6):H1911-H1917.
48. Moses S, Dreja K, Lindqvist A, Lovdahl C, Hellstrand P, Hultgardh-Nilsson A: **Smooth muscle cell response to mechanical injury involves intracellular calcium release and ERK1/ERK2 phosphorylation.** *Experimental Cell Research* 2001, **269**(1):88-96.
49. Martin PEM, Wall C, Griffith TM: **Effects of connexin-mimetic peptides on gap junction functionality and connexin expression in cultured vascular cells.** *British Journal of Pharmacology* 2005, **144**(5):617-627.
50. Okamoto T, Akiyama M, Takeda M, Gabazza EC, Hayashi T, Suzuki K: **Connexin32 is expressed in vascular endothelial cells and participates in gap-junction intercellular communication.** *Biochemical and Biophysical Research Communications* 2009, **382**(2):264-268.
51. Evans WH, Boitano S: **Connexin mimetic peptides: specific inhibitors of gap-junctional intercellular communication.** *Biochemical Society Transactions* 2001, **29**.
52. Tang EHC, Vanhoutte PM: **Gap junction inhibitors reduce endothelium-dependent contractions in the aorta of spontaneously hypertensive rats.** *Journal of Pharmacology and Experimental Therapeutics* 2008, **327**(1):148-153.
53. Brisset AC, Isakson BE, Kwak BR: **Connexins in Vascular Physiology and Pathology.** *Antioxidants & Redox Signaling* 2009, **11**(2):267-282.
54. Kruger O, Beny JL, Chabaud F, Traub O, Theis M, Brix K, Kirchhoff S, Willecke K: **Altered dye diffusion and upregulation of connexin37 in mouse aortic endothelium deficient in connexin40.** *Journal of Vascular Research* 2002, **39**(2):160-172.
55. Simon AM, McWhorter AR: **Decreased intercellular dye-transfer and downregulation of non-ablated connexins in aortic endothelium deficient in connexin37 or connexin40.** *Journal of Cell Science* 2003, **116**(11):2223-2236.
56. Isakson BE, Damon DN, Day KH, Liao YB, Duling BR: **Connexin40 and connexin43 in mouse aortic endothelium: evidence for coordinated regulation.** *American Journal of Physiology-Heart and Circulatory Physiology* 2006, **290**(3):H1199-H1205.
57. Isshiki M, Anderson RGW: **Function of caveolae in Ca²⁺ entry and Ca²⁺-dependent signal transduction.** *Traffic* 2003, **4**(11):717-723.
58. Isshiki M, Ying YS, Fujita T, Anderson RGW: **A molecular sensor detects signal transduction from caveolae in living cells.** *Journal of Biological Chemistry* 2002, **277**(45):43389-43398.
59. Munaron L: **Intracellular calcium, endothelial cells and angiogenesis.** *Recent Patents on Anti-Cancer Drug Discovery* 2006, **1**(1):105-119.
60. Fischell TA, Derby G, Tse TM, Stadius ML: **Coronary-artery vasoconstriction routinely occurs after percutaneous trans-luminal coronary angioplasty - a quantitative arteriographic analysis.** *Circulation* 1988, **78**(6):1323-1334.
61. Yoder MC: **Human endothelial progenitor cells.** *Cold Spring Harbor perspectives in medicine* 2012, **2**(7).
62. Schwartz SM, Benditt EP: **Clustering of replicating cells in aortic endothelium.** *Proceedings of the National Academy of Sciences of the United States of America* 1976, **73**(2):651-653.
63. Parthasarathi K, Ichimura H, Monma E, Lindert J, Quadri S, Issekutz A, Bhattacharya J: **Connexin 43 mediates spread of Ca²⁺-dependent proinflammatory responses in lung capillaries.** *Journal of Clinical Investigation* 2006, **116**(8):2193-2200.
64. Schalper KA, Sanchez HA, Lee SC, Altenberg GA, Nathanson MH, Saez JC: **Connexin 43 hemichannels mediate the Ca²⁺ influx induced by extracellular alkalization.** *American Journal of Physiology-Cell Physiology* 2010, **299**(6):C1504-C1515.
65. Alonso F, Boittin F-X, Beny J-L, Haeffliger J-A: **Loss of connexin40 is associated with decreased endothelium-dependent relaxations and eNOS levels in the mouse aorta.** *American Journal of Physiology-Heart and Circulatory Physiology* 2010, **299**(5):H1365-H1373.
66. Straub AC, Billaud M, Johnstone SR, Best AK, Yemen S, Dwyer ST, Looft-Wilson R, Lysiak JJ, Gaston B, Palmer L, et al: **Compartmentalized Connexin 43 S-Nitrosylation/Denitrosylation Regulates Heterocellular Communication in the Vessel Wall.** *Arteriosclerosis Thrombosis and Vascular Biology* 2011, **31**(2):399-U353.
67. Pfenniger A, Derouette J-P, Verma V, Lin X, Foglia B, Coombs W, Roth I, Satta N, Dunoyer-Geindre S, Sorgen P, et al: **Gap Junction Protein Cx37 Interacts With Endothelial Nitric Oxide Synthase in Endothelial Cells.** *Arteriosclerosis Thrombosis and Vascular Biology* 2010, **30**(4):827-U445.
68. Rath G, Dessy C, Feron O: **Caveolae, caveolin and control of vascular tone: nitric oxide (no) and endothelium derived hyperpolarizing factor (edhf) regulation.** *Journal of Physiology and Pharmacology* 2009, **60**:105-109.
69. Schubert AL, Schubert W, Spray DC, Lisanti MP: **Connexin family members target to lipid raft domains and interact with caveolin-1.** *Biochemistry* 2002, **41**(18):5754-5764.
70. Saliez J, Bouzin C, Rath G, Ghisdal P, Desjardins F, Rezzani R, Rodella LF, Vriens J, Nilius B, Feron O, et al: **Role of caveolar compartmentation in endothelium-derived hyperpolarizing factor-mediated relaxation - Ca²⁺ signals and gap junction function are regulated by caveolin in endothelial cells.** *Circulation* 2008, **117**(8):1065-1074.
71. Weber C: **Platelets and chemokines in atherosclerosis - Partners in crime.** *Circulation Research* 2005, **96**(6):612-616.
72. Lee PC, Salyapongse AN, Bragdon GA, Shears LL, Watkins SC, Edington HDJ, Billiar TR: **Impaired wound healing and angiogenesis in eNOS-deficient mice.** *American Journal of Physiology-Heart and Circulatory Physiology* 1999, **277**(4):H1600-H1608.
73. Guo JP, Panday MM, Consigny PC, Lefer AM: **Mechanisms of vascular preservation by a novel no donor following rat carotid-artery intimal injury.** *American Journal of Physiology-Heart and Circulatory Physiology* 1995, **269**(3):H1122-H1131.
74. Murohara T, Asahara T, Silver M, Bauters C, Masuda H, Kalka C, Kearney M, Chen DH, Chen DF, Symes JF, et al: **Nitric oxide synthase modulates angiogenesis in response to tissue ischemia.** *Journal of Clinical Investigation* 1998, **101**(11):2567-2578.

75. Amano K, Matsubara H, Iba O, Okigaki M, Fujiyama S, Imada T, Kojima H, Nozawa Y, Kawashima S, Yokoyama M, et al: **Enhancement of ischemia-induced angiogenesis by eNOS overexpression.** *Hypertension* 2003, **41**(1):156-162.
76. Dimmeler S: **Regulation of Bone Marrow-Derived Vascular Progenitor Cell Mobilization and Maintenance.** *Arteriosclerosis Thrombosis and Vascular Biology* 2010, **30**(6).
77. Isakson BE, Ramos SI, Duling BR: **Ca²⁺ and inositol 1,4,5-trisphosphate-mediated signaling across the myoendothelial junction.** *Circulation Research* 2007, **100**(2):246-254.
78. Sanchez HA, Orellana JA, Verselis VK, Saez JC: **Metabolic inhibition increases activity of connexin-32 hemichannels permeable to Ca²⁺ in transfected HeLa cells.** *American Journal of Physiology-Cell Physiology* 2009, **297**(3):C665-C678.
79. Sanchez HA, Mese G, Srinivas M, White TW, Verselis VK: **Differentially altered Ca(2+) regulation and Ca(2+) permeability in Cx26 hemichannels formed by the A40V and G45E mutations that cause keratitis ichthyosis deafness syndrome.** *Journal of General Physiology* 2010, **136**(1):47-62.
80. Schalper KA, Palacios-Prado N, Retamal MA, Shoji KF, Martinez AD, Saez JC: **Connexin hemichannel composition determines the FGF-1-induced membrane permeability and free Ca²⁺ (i) responses.** *Molecular Biology of the Cell* 2008, **19**(8):3501-3513.
81. Sorensen CM, Salomonsson M, Braunstein TH, Nielsen MS, Holstein-Rathlou N-H: **Connexin mimetic peptides fail to inhibit vascular conducted calcium responses in renal arterioles.** *American Journal of Physiology-Regulatory Integrative and Comparative Physiology* 2008, **295**(3):R840-R847.
82. Shestopalov VI, Panchin Y: **Pannexins and gap junction protein diversity.** *Cellular and Molecular Life Sciences* 2008, **65**(3):376-394.
83. Gossman DG, Zhao H-B: **Hemichannel-Mediated Inositol 1,4,5-Trisphosphate (IP3) Release in the Cochlea: A Novel Mechanism of IP3 Intercellular Signaling.** *Cell Communication and Adhesion* 2008, **15**(4):305-315.
84. Varu VN, Tsihlis ND, Kibbe MR: **Nitric Oxide-Releasing Prosthetic Materials.** *Vascular and Endovascular Surgery* 2009, **43**(2):121-131.
85. De Vuyst E, Boengler K, Antoons G, Sipido KR, Schulz R, Leybaert L: **Pharmacological modulation of connexin-formed channels in cardiac pathophysiology.** *British Journal of Pharmacology* 2011, **163**(3).
86. Salameh A, Dhein S: **Pharmacology of Gap junctions. New pharmacological targets for treatment of arrhythmia, seizure and cancer?** *Biochimica Et Biophysica Acta-Biomembranes* 2005, **1719**(1-2).

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